The Halictine Bees of Sri Lanka and the Vicinity. II. Nesohalictus (Hymenoptera: Halictidae)

Shôichi F. Sakagami

Zoological Section, Institute of Low Temperature Science Hokkaido University, Sapporo 060, Japan

ABSTRACT—Redescriptions of two halictine bee species, Lasioglossum (Nesohalictus) serenum (Cameron) from Sri Lanka and India and L. (N.) halictoides (Smith) from Insular Malesia, and taxonomic notes on Nesohalictus characterized by specialized glossa and femoral scopa.

INTRODUCTION

Nesohalictus Crawford [1] is an Indomalayan subgenus of the large halictine genus Lasioglossum Curtis and is distinguished from congeneric subgenera by the unusually long glossa (Figs. 1, 7A, B) and sparse and simplified femoral scopa (Fig. 2D) [2]. This subgenus contains three species [3, 4]. In the present paper, L(N.) serenum (Cameron) from Sri Lanka and India is compared with L. (N.) halictoides (Smith), the type species of Nesohalictus, together with some taxonomic notes on the subgenus.

RESULTS

Features Common to L. serenum and L. halictoides

Female: *Coloration* Non-metallic and basically black; tergal margins not much paler. Mandible dark brown, apically chestnut brown.

Pilosity Hairs pale except some brownish hairs on mesoscutum and mesoscutellum, and simple, erect, dark hairs on metasomal terga. Vestiture moderately dense, not hiding surface except tomentum on gena along outer orbit, pronotum and basal tergal fasciae. *Head* Vertex with long



FIG. 1. L. serenum (2), with the mouth parts extended.

Accepted June 28, 1990 Received June 17, 1990 (275 µm), erect, plumose hairs, mixed with denser, shorter (30-50 µm), simple hairs, the latter also on ocellocular and circumocellar areas and on frons above. Long hairs around antenna (300 µm) becoming shorter (100 µm), semierect on paraocular area (Fig. 2A) mixed with short, appressed, tomental hairs but not completely hiding surface; on paraocular area below gradually changing to plumose, semierect hairs $(\pm 75 \,\mu m)$. Hairs on supraclypeus rather sparse, plumose, semierect $(75-125 \ \mu m)$; on clypeus 175-225 μm , poorly plumose and appressed, apical bristles to $375 \,\mu m$. Gena tomented with short (30 μ m), plumose hairs, denser along outer orbit; gradually sparser postward admixed with erect, plumose hairs (175 μ m), the latter sparser and longer $(375 \,\mu\text{m})$ toward Mesosoma Pronotum hypostoma. densely tomented, anteriorly with erect, plumose hairs (200 µm). Mesoscutum with erect to semierect, moderately dense hairs (150–200 μ m); underhairs represented only by sparse, narrow tomentum along lateral margin. Mesoscutellum similar, posterior fringe attaining 500 µm. Metanotal tomentum dense, admixed with moderately dense, plumose, erect hairs. Propodeal dorsum glabrous except triangular, sparse, tomental patch on posterolateral area (Fig. 3). Mesopleuron (above 175 μ m, below 250 μ m), and propodeal side and declivity (250 µm) with moderately dense, erect, plumose hairs; underhairs tomented on metapleuron and propodeal side, virtually absent on mesopleuron. Tegula anteriorly with dense, semierect, plumose hairs (100 μ m); gradually shorter, sparser and simple postward; posterolateral two thirds glabrous. Fore trochanter and femur below with plumose, relatively sparse hairs (300-375 μ m). Mid leg with trochanter and femoral hairs relatively short (300 and 250 µm; in other Lasioglossum, e.g. L. duplex, 500 and 375 µm). Trochanter and femoral scopa of hind leg distinctly sparse, and rather sparsely branched (Fig. 1D vs C). Metasoma Tergum 1 (T_1) on basal slope with dense, erect, plumose hairs (medially 250 µm, laterally to $300 \,\mu m$), above moderately tomented, disc with sparse, simple, pale hairs $(20-25 \,\mu m)$, lateral fringe (to 150 µm) plumose. T2-T4 basally with tomental fasciae. T₂ posteriorly homogeneously with yellowish, fine, simple hairs (25-40 μ m), dense but invisible from some direction, sparsely admixed with stouter, darker, semierect hairs (50 μ m). T₃-T₄ similar but posterior hairs gradually longer (to 125 μ m) and darker; with semierect, dark hairs dense, some ones poorly and sparsely branched (to 250 μ m); whitish lateral fringe to 400 μ m on T₄. T₅₋₆ predominated with short, dark hairs except whitish lateral fringe. Ventrolateral areas of terga with hairs sparse, long (max. 500 μ m), simple but some ones poorly plumose. Sternal hairs sparse, simple (max. 750 μ m).

Structure. Head distinctly narrower than mesosoma, moderately elongate; inner orbits below rather straightforward convergent; outer orbits moderately rounded, convergent below (Fig. 2A). Eye with fine, very sparse setae, seen glabrous. Vertex seen frontally gently convex; lateral ocellus not attaining summit, lower margin on supraorbital line; ocellocular area with fine (ϕ 15–12 μ m) and shallow punctures (PP); interspaces (IS) not linear but narrower than PP, dully shining; ocellar and postocellar areas similar, on the latter PP tending to form transverse rows; ocellar area gently raised, posteriorly mildly depressed. Occiput carinate. Frons flat, seen microareolate, with IS linear and areolae small ($\phi 20 \,\mu m$ or less) and acute, forming oblique-longitudinal rows; frontal carina distinct, above replaced by fine, linear sulcus attaining near mid ocellus. Supraclypeus and paraocular area finely meshlike tessellate, dully shining with obscure, shallow PP ($\phi 25-35 \mu m$, IS/ $PP\phi = 1.0$ or more); supraclypeus gently raised above. Epistomal angle acute, epistomal lobe developed (Fig. 2A). Clypeus rather flat, the part below suborbital line longer than half the clypeal length; above sculptured as on supraclypeus but IS =1.0-3.0; below obscurely, longitudinally undulate, smooth and shining, PP ϕ 25-40 μ m, often elongate; apically transversely depressed, lateral tooth mild but distinct. Gena moderately wide, above not swollen, sculptured as on ocellocular area but PP finer. Hypostoma mat with dense striation. Malar space linear. Labrum (Fig. 2E, F) with large, transverse-rectangular, flat, basal tubercle; apical part triangular with distinct median keel, subapically tuberculate. Mandible bidentate (Fig. 2A). Glossa extremely long (Figs. 1, 7A,B), maxillary and labial palpi normal (6- and

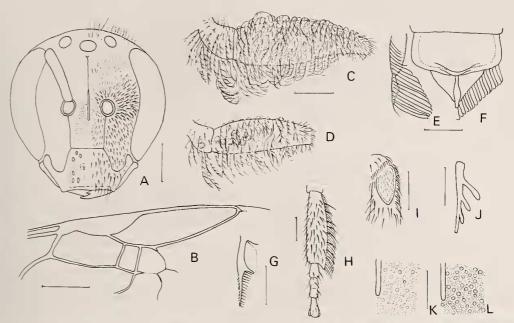


FIG. 2. Female characters of L. serenum (A, B, D-K), L. duplex (C) and L. halictoides (L). A, face seen frontally; B, fore wing; C, D, femoral scopa; E, F, lateral and dorsal view of labrum; G, strigilis (antenna cleaner); H, fore tarsi; I, basitibial plate; J, inner hind tibial spur; K, L, mesoscutal sculpture. Scale=0.5 mm in A-D, 0.25 mm in E-L.

4-segmented, ratios 5/7/7/8/8/8 and 10/5/6/6 in *L. serenum* (Fig. 7A, B). Scape attaining mid ocellus.

Pronotum dorsally concave; lateral angle very obtuse, inconspicuously angulate seen both frontally and dorsally; dorsal ridge carinate; lateral ridge vestigial; lateral surface and lobe virtually unseparated. Mesoscutum anteriorly neither projecting nor bilobed, roundly truncate; declivity not forming differentiated lip; median line not weakened throughout; parapsidal line distinct; disc coriaceous and dull, with obscure PP ($\phi = 520 \mu m$) (Fig. 2K, L), denser, finer and more distinct near posterior margin. Mesoscutellum flat, medially not depressed, sculptured as on mesoscutum. Mesopleuron strongly reticulate, rather irregularly above and anteriorly; the rest forming dense, transverse carinulae. Propodeal dorsum (Fig. 3) mildly sloping, subapically limited by mild, crescent ridge; basally coriaceous, dull with rather sparse, strong rugae, either longitudinal and radiated laterally or irregular, often anastomosing; postward not extending beyond crescent ridge; lateral and posterior margins strongly carinate

though weakened medially where confluent with crescent ridge; posterolateral angle acute but not pointed. Tegula dully shining with superficial tessellation.

Fore basitarsal comb entire, accompanied with long hairs nearby (Fig. 2H). Strigilis of common *Lasioglossum* type (Fig. 2G); malus as long as vellum, outer margin denticulate; vellum with inner margin straight. Mid and hind legs normal; basitibial plate elliptical, apically rather pointed (Fig. 2I). Inner hind tibial spur (Fig. 2J) with 3-4 rather long flat teeth, the most apical one often small. Both hind tibia and basitarsus slender, the latter parallel-sided, the ratio tibia, basi- and distitarsi 42:23:23. Marginal cell apically apart from wing margin; tc 3 distinctly reduced but tc 2 not (Fig. 2B).

Metasoma elongate oval. Tergum 1 not pedunculate, boundary between basal slope and disc rounded; basal slope distinctly but superficially tessellate, dully shining with ϕ PP 12–20 μ m and IS 3.0 or more; disc similarly sculptured but tending to lineolate with PP finer (ϕ 10–12 μ m), IS 1.0–2.0; sparse (2.0–3.0) on very mild boss; mar-

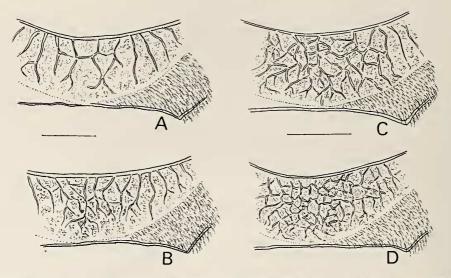


Fig. 3. Propodeal dorsum of females of *L. serenum* (A, B) and *L. halictoides* (C, D). A, B, specimens with ridges sparse and dense; C, D, specimens from Kota Kinabalu (Borneo) and Manado (Sulawesi). Scale=0.25 mm.

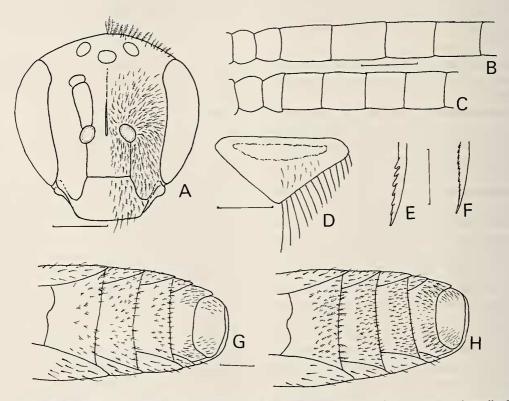


FIG. 4. Male characters of L. serenum (A, C, D, F, H) and L. halictoides (B, E, G). A, face seen frontally; B, C, basal flagellomeres; D, labrum seen dorsally; E, F, inner hind tibial spur, G, H; sternal pilosity. Scale =0.5 mm in A, G, H; 0.25 mm in others.

ginal area mildly depressed only behind boss; PP sparser than on disc. T_2 etc. similar but tessellation more conspicuous and PP gradually coarser on posterior terga though always weak and ϕ never exceeding 25 μ m; boss and marginal area more clearly differentiate even though still inconspicuously. Sterna normal, densely lineolate, post-gradular area coarsely granulate.

Male Coloration as in female, clypeus and legs without pale markings. *Pilosity* as in female: Plumose hairs on paraocular area denser and more appressed. Hairs on legs moderately sparse, on tibiae and basitarsi relatively long; hairs on fore leg attaining 250 μ m, on mid femur below 175 μ m, mid basitarsus below 200 μ m, hind femur 200 μ m, basitarsus 250 μ m, all poorly plumose and rather erect. Sterna (Fig. 4G, H) with rather sparse homogeneous, semierect and poorly plumose hairs.

Structure Except metasomal terminalia similar

to female but (1) eye more swollen (Figs. 2A, 4A), (2) scape shorter, not attaining mid ocellus (Figs. 2A, 4A, 6), (3) flagellomeres longer (FIg. 6), (4) vertex more raised (Figs. 2A, 4A), (5) labrum (Fig. 4D) triangular, flat, basally mildly raised transversely, apically mildly pointed, (6) mandible edentate, (7) clypeal tooth obsolete, (8) legs of normal male type, hind basitarsus apically more convergent, (9) inner hind tibial spur (Fig. 4E, F) with appressed denticles, (10) hind distitarsi slender; ratio hind tibia, hind basi - and distitarsi 38/ 20/20.

Metasomal sternum 5 (S₅, Fig. 5A, B) with basal margin widely incurved, apically deeply emarginate, lateral process sparsely haired, gradulus transverse, curved laterally. S₆ basally deeply incurved, apodemal lobe long, gradulus transverse, evanescent laterally (Fig. 5C, D). S₇ apically projecting. S₈ medially very elongate, apically tapering and finely haired. (Fig. 5E, F). Gonobase long, para-

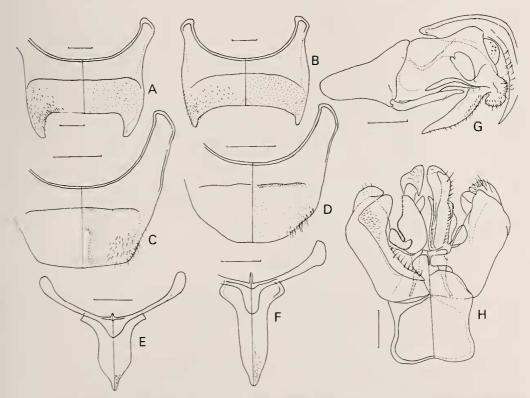


FIG. 5. Male terminalia of L. serenum (B, D, E, G, H) and L. halictoïdes (A, C, F). A, B, Sternum 5 (hairs shown in left half, sculpture in right half), C, D, S₆; E, F, S_{7.8}; G, H, genitalia seen laterally (G), ventrally (H, left) and dorsally (H, right). Scale=0.33 mm in C, D; 0.25 mm in others.

llel-sided (Fig. 5G, H); gonocoxite (Fig. 5H) not continuing gonobasal outline, outer margin not outcurved but angulate, about two times longer than wide; gonostylus (Fig. 5G) short and rounded, sparsely haired; retrose lobe (Fig. 5G, H) very long, slender, apically pointed, with fine short hairs basally and long erect hairs apically. both sexes of the two species. On the average, *L.* serenum is smaller in most characters but the values are very similar between the two species. By non-overlap of SD, only the following characters show the significant difference (indicated with arrows, s=serenum, h=halictoides, $\bar{x}\pm$ SD in parentheses (40 units=1 mm, n=4 in WD, Sm2L, Sm3L in $h \diamondsuit$, n=5 in all others). Female: CAL (s 35.1 ± 1.3 , $h 38.4\pm2.7$), IAD ($s 8.3\pm0.45$, $h 9.4\pm$ 0.5); Male: McL ($s 37.0\pm1.4$, $h 41.6\pm1.4$), Sm2L ($s 5.0\pm1.6$, $h 8.6\pm1.2$), CAL ($s 32.0\pm1.4$, h 36.0

Morphometric Comparison

Figure 6 compares main metric characters of

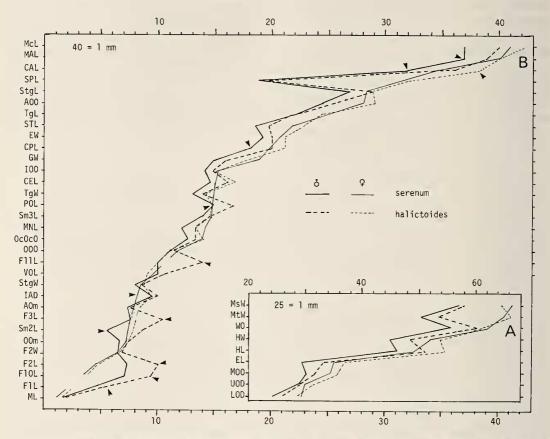


FIG. 6. Comparison of male and female metric characters in L. serenum and L. halictoides, arranged in both A (25 units=1 mm) and B (40 units=1 mm) in the descending order in L. serenum female. L, W, D=length, width, distance. A: MsW, MtW (meso- and metasoinal W), WD (Wing diagonal=D between M-Cu bifurcation and inner tip of marginal cell), HW, HL (head W and L), EL (eye L), MOD, UOD, LOD (maximum, upper, and lower interorbital D). B: McL (marginal cell L), MAL (L betwen marginal cell tip and wing tip), CAL (clypealveolar D), SPL (scape L), StgL (pterostigma L), AOD (alveocellar D), TgL (tegula L), STL (scutellulm L), EW (eye W, seen laterally), CPL (clypeus L), GW (gena W, seen laterally), IOD (interocellar D), CEL (L of apical clypeal part exceeding lower orbital line), TgW (tegula W), PDL (propodeal dorsum L), Sm3L, Sm2L (submarginal cell 2, 3 L), MNL (metanotum L), OCOCD (ocelloccipital D), OOD (ocellocular D), FnL, W (flagellomere n L, W), VOL (verticorbital L=tangential L between summit of vertex and supraorbital line), StgW (pterostigma W), IAD (interalveolar D), ADm, ODm (alveolus and mid ocellus diameter), ML (malar L).

174

 ± 1.9), CPL (s 18.2 ± 0.8 , h 20.2 ± 1.2), PDL (s 14.9 ± 0.8 , h 16.8 ± 0.8), F11L (s 10.0 ± 0.6 , h 14.0 ± 0.3), F3L (s 7.7 ± 0.52 , h 10.5 ± 0.3), F2L (s 7.2 ± 0.5 , h 10.1 ± 0.4), F10L (s 7.0 ± 0.3 , h 9.4 ± 0.5), F1L (s 4.2 ± 0.3 , h 5.2 ± 0.2). Male flagellomeres are distinctly longer in L. halictoides (Fig. 4B, C).

Some important ratios are also not much different between the two species HW/MsW/MtW (φ both *s*, *h* 1/1.28/1/25; \Diamond *s* 1/1.27/1.12, *h* 1/1.19/1.10), HW/HL (φ *s* 1/0.99, *h* 1/1.01; \Diamond *s* 1/1.03, *h* 1/1.05), UOD/LOD (φ *s* 1/0.95, *h* 1/0.98; \Diamond *s* 1/0.90, *h* 1/0.89), CPL/CAL/CEL (φ *s* 1/1.80/0.77 *h* 1/1.81/0.80; \Diamond both *s*, *h* 1/1.76/0.80), IOD/OOD (φ *s* 1/0.75, *h* 1/0.73; \Diamond both *s*, *h* 1/1.079), EW/GW (φ *s* 1/0.90, *h* 1/0.85; \Diamond both *s*, *h* 1/0.78), SCL/MNL/PDL (φ *s* 1/0.64/0.67, *h* 1/0.59/0.70; \Diamond *s* 1/0.65/0.81, *h* 1/0.68/0.85), HW/WD (φ *s* 1/1.19, *h* 1/1.14; \Diamond *s* 1/1.24, *h* 1/1.26).

The most conspicuous feature of Nesohalictus, the length of elongate glossa, was measured only in several specimens (length of glossa/ratio length of glossa to wing diagonal): ♀, serenum (1.5 mm/ 0.25, 1.7 mm/0.30), halictoides (2.0 mm/0.38, 2.6 mm/0.43, 3.2 mm/0.61*); \updownarrow , serenum (1.3 mm/ 0.25, 1.4 mm/0.25, 1.5 mm/0.25, 1.6 mm/0.32*, 2.0 mm/0.38), halictoides (2.0 mm/0.38, 2.3 mm/ 0.42, 2.5 mm/0.45, 2.9 mm/0.49, 3.6 mm/0.65*). Mouth parts are extended foreword in asterisked specimens (Fig. 7B) and flexed in others. In the former position the glossa is longer possibly because the basal part is fully extended. From all obtained results, it is concluded that L. halictoides has the glossa longer than L. serenum in both the absolute length as well as the length relative to the wing length.

Structural Comparison

Female (1) Color generally paler in s (serenum), especially tegula pale brown against dark to blackish brown in h (halictoides). Veins pale brown in s, brown in h; pterostigma and subcosta brown in s, chestnut to dark brown in h. (2) Tergum 1 (and often also T_2 basally) pale reddish brown in s, homogeneously dark in h. (3) Hairs paler, usually whitish in s, more yellowish in h. (4) Basal fasciae of T_2 - T_4 wider and continuous in s, narrower and often medially interrupted in h. (5) Mesoscutal and -scutellar PP rather ill-defined and sparser, IS = 1.5-3.0 of ϕ PP in s (Fig. 2K), more distinct and denser, often ϕ PP>IS even on scutal disc medially in h (Fig. 2L). (6) Rugae of propodeal dorsum on the average sparser and seldom attaining crescent subapical ridge in s (Fig. 3A, B), denser and often attaining ridge in h (Fig. 3C, D). (7) Tegular sculpture more superficial in s than in h.

Male (1) Coloration as in female but terga of soften darker, ranging from pale brown to blackish. (2) Sternal hairs of s (Fig. 4H) denser, not confined to apical half of each sternum, more distinctly plumose and, semiappressed in s (Fig. 4H); sparser, confined to apical half and rather erect, only appressed marginally in h (Fig. 4G). (3) Sterna with more distinct tessellation and duller in s, more superficially tessellate and shiner in h. (4) Denticles of hind inner tibial spur finer and homogeneous in s (Fig. 4F), stronger and middle ones longest (Fig. 4E) in h. (5) Posterior margin of metasomal sternum 5 (S_5) gently incurved in s, transverse in h (Fig. 5B, A). (6) S₆ with apodemal lobe shorter and apical margin only medially truncate in s (Fig. 5D), lobe longer and apex more widely truncate in h (Fig. 5C). (7) Median lobe of S_7 shorter and apically acutely tapering in s, longer and mildly tapering in h. (8) Median part of S_8 triangular in s, slender and elongate in h (Fig. 5E, F). (9) Gonostylus seen dorsally more triangular in s, rather rounded in h.

Synonymy and Distribution

Lasioglossum (Nesohalictus) halictoides (Smith)

Andrena (nec Nomia as cited by Blüthgen 1931) halictoides Smith, 1859, J. Proc. Linn. Soc. London, Zool., 3 (1858): 6, $\stackrel{\circ}{\rightarrow}$ (Celebes).

Halictus halictoides, Cockerell, 1922, Ann. Mag. Nat. Hist., (9) 9: 662; Blüthgen, 1930, Mitt. deuts. entom. Gesell., 75 (syn. *H. biroi*).

H. biroi Friese, 1909, Ann. Mus. Nat Hunger., 7: 188, Blüthgen, 1926, Zool. Jb., Syst., 51: 541 (Key to allied spp. $\stackrel{\circ}{+}$; syn. *carinatifrons*, etc.), (New Guinea).

H. carinatifrons Strand, 1910, Berlin. entom. Zts., 54, (1909): 196, ♦, ♀: Blüthgen, 1922, Deuts. entom. Zts., 53, 54 (synn.); 1926, Zool.

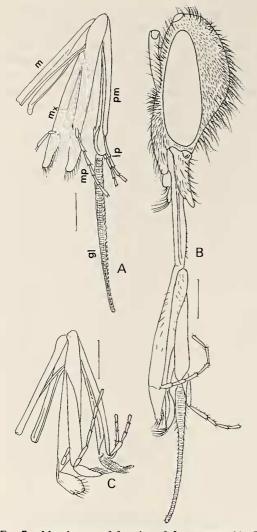


FIG. 7. Mouthparts of females of *L. serenum* (A, B) and *L. duplex* (C). A, C, mentum and prementum, flexed; g=glossa, mx=maxilla, mp, lp=maxillary and labial palpi, m=mentum, pm=prementum. B, mentum and prementum extended.

Jb., Syst., 51: 541 (Taiwan).

H. heymonsii Strand, 1910, Berlin. entom. Zts., 54 (1909): 207, ☆; Blüthgen, 1922, Deuts. entom. Zts., 53 (syn. carinatifrons), (Taiwan).

H. blepharophorus Strand, 1913, Supplm. Entom., 2: 28 \updownarrow (nec $\updownarrow = micado$ Strand, 1910 = *cattulus* Vachal, 1894, $\varUpsilon = vagans$ Smith, 1857, \varUpsilon); Blüthgen, 1923, Deuts. entom. Zts.: 242 (= *ceylonicus* Strand 1910, \updownarrow); 1926, Zool. Jb., Syst., 51: 541 (Sri Lanka).

H. taihorinis var. anpingensis Strand, 1914, Arch. Naturg. 79A: 151, \Im ; Blüthgen, 1923, Deuts. entom. Zts.: 241 (=*ceylonicus* Strand, 1910 \Im); 1926, Zool. Jb., Syst., 51: 542.

H. lativentris Friese, 1914 (nec Schenck, 1853), Tijdschr. Entom., 57: 22, 2, 3 (Java); Blüthgen, 1925, Deuts. entom. Zts.: 400 (=*carinatifrons* Strand, 1910); 1926, Zool. JB., Syst., 51: 542.

H. (*Nesohalictus*) robbii Crawford, 1910, Proc. U. S. Nat. Mus., 38: 120, \uparrow , \updownarrow (Philippines); Blüthgen, 1925 Deuts. entom. Zts., 1925: 415 (= *carinatifrons* Strand, 1910); 1931, Zool. Jb., Syst., 61: 300.

Nesohalictus robbii, Cockerell, 1919, Phil. J. Sci., 15: 269.

Lasioglossum (Nesohalictus) biroi, Michener, 1965, Bull. Amer. Mus. Nat. Hist., 130: 174.

Blüthgen [3, 4] synonymized various names with L. halictoides. Cockerell [5] is skeptical for this lumping. Here the former treatment is adopted although a subspecific differentiation is likely to occur in this species widely distributed in various islands. Blüthgen [3] synonymized Halictus blepharophorus Strand from Sri Lanka with L. halictoides but later did not mention the occurrence of L. halictoides in Sri Lanka. It is likely that H. blepharophorus is synonymous with L. serenum from Sri Lanka, not with L. halictoides. Critical comparison of the type specimen of H. blepharophorus with both L. serenum and L. halictoides is necessary.

Specimens examined: Sabah (new record) Jesselton (now Kota Kinabalu), $3 \Leftrightarrow 10$ 1965 (one \Leftrightarrow with an emergence hole of stylops between terga 3-4), $2 \Leftrightarrow 7$ ix 1966; Sarawak (new record) Kuching $2 \Leftrightarrow 8$ ix 1966; Manado (previously Menado), Sulawesi, $1 \Leftrightarrow$, 1 iii 1984; Krakatau (new record): Anak Krakatau, $1 \Leftrightarrow$, 29 vii 1982; Rakata, $1 \Leftrightarrow$, 30-31 viii, 1984.

Distribution: Taiwan, Philippines, Java, Borneo, Sulawesi, Krakatau, New Guinea, ? Sri Lanka.

Lasioglossum (Nesohalictus) serenum (Cameron) comb. nov.

Halictus serenus Cameron, 1897, Mem. Manchester Soc., 41: 97, ↑ (India); Blüthgen, 1930, Mitt. deuts. entom. Gesell., 1930: 76 (=strandiellus Cockerell); 1931, Zool. Jb., Syst., 61: 300.

H. deesanus Cameron, 1908, J. Bombay Nat. Hist. Soc., 13: 309, 2 (India); Blüthgen, 1931, Mitt. deuts. entom. Gesell., 1931, 76 (=*H. serenus* Cameron).

H. ceylonicus Strand, 1910 (nec Cameron, 1902 =alphenum Cameron, 1899), Berlin. entom. Zts., 54 (1909): 187, ↑; Blüthgen, 1922, Deuts. entom. Zts.,: 53 (=carinatifrons Strand, 1910); 1925, Deuts. entom. Zts.,: 385 (=strandiellus Cockerell 1911).

H. strandiellus Cockerell, 1911, Ann. Mag. Nat. Hist. (8) 8: 192 (=n. n. for *ceylonicus* Strand, nec Cameron); Blüthgen, 1925, Deuts. entom. Zts.,: 385; 1926, Zool. Jb. Syst., 51: 541.

H. hornianus Strand, 1913, Arch. Naturg., 79,
A, 2: 138, [↑]; Blüthgen, 1925, Deuts. entom. Zts.:
399 (=strandiellus Cockerell); Blüthgen, 1926,
Zool. Jb., Syst. 51: 542.

Specimens examined: Sri Lanka Col. Dist., Colombo, Museum Gardens, 1 3 18 i 1977; Ham. Dist., Palatupana tank, 1♀ 21-22 vi 1978, Yala, Palatupana, 1 & 21-22 vi 1978; Kan. Dist., Kandy, Udawattakele Sanctuary, 2100 ft, 1 1-17 ix 1976, 1600 ft, 1 118-21 i 1977; Man. Dist., Cashew Corp., Ma Villu, 1♀ 17-21 ii 1979; Mon. Dist., Angunakalapelessa (Malaise trap), 1♀ 2☆ 17-19 vi 1978; Pol. Dist., 25 mi SE Pelonnaruwa, 1 ☆, 10 vi 1975; Put. Dist., Dedunioya, 1 ♀ 5 iii 1958; Vav. Dist., Parayanalankulam Irrigation Canal, 25 mi NW Medawachchiya, 100 ft, 1♀ 20-25 iii 1970. India Kerala: Walayer, subtropical monsoon forest, Ipomea, 1º, 29 i 1978; Tamil Nadu: Coimbatore 3 𝔅, 5-10 xii 1978, Madras, City Park, 13, 19 viii 1975.

Distribution: Sri Lanka, India.

Taxonomic and Bionomic Notes

Nesohalictus is closely allied to Ctenonomia, the large palaeotropic subgenus of Lasioglossum [6]. Apart from its long glossa and simplified femoral scopa, Nesohalictus could be regarded as a specialized species group of Ctenonomia. Among the species groups of Ctenonomia, the carinatum group is similar to Nesohalictus group by the carinate occiput (previously the occiput of this group was erroneously described as "carinate or not", 6), and lateral and posterior margins of propodeal dorsum continuously carinate, but differs by posterolateral corner of propodeal dorsum not glabrous but haired as in the *vagans* group whose occiput is not carinate.

The peculiar fore basitarsal comb and hind femoral scopa were assumed as adaptations to collect coarse pollen such as of Hibiscus [2]. The presence of such coarse pollen within scopa in one female of L. serenum (Fig. 2D) from Sri Lanka and one female of L. halictoides from Krakatau favors the above assumption. On the other hand, the flower preference of Nesohalictus for nectar intake is still unknown. Blüthgen [3] mentioned that glossa of L. serenum is about as long as that of L. halictoides but actually shorter as aforementioned. The third known species, L. (N.) goluratum (Blüthgen, 3) from Burma and Penang has the distinctly shorter glossa. The three species seem to form a series of the prolongation of glossa (halictoides > serenum > goluratum).

Nothing is known on the nest architecture, life cycle and social pattern of the three *Nesohalictus* species. Clarification of their bionomics by residential naturalists is requested.

ACKNOWLEDGMENTS

I thank all colleagues and friends who collected examined specimens or put them at my disposal for studies, particularly Dr. K. V. Krombein (Department of Entomology, Smithsonian Institution, Washington, D. C.) and Prof. S. Takagi and Dr. T. Kumata (Entomological Institute, Hokkaido University, Sapporo). This paper is a part of "Biosystematic studies of the Insects of Sri Lanka" directed by Dr. Karl V. Krombein, Smithsonian Institution, Washington, D. C. and "Research Trips for Forest and Agricultural Insects in the Subcontinent of India JICT: (Hokkaido University, University of Calcutta and Zoological Survey of India Joint Project) Scientific Report Nr. 44".

REFERENCES

- Crawford, J. C. (1910) New Hymenoptera from the Philippine Islands. Proc. U. S. Nat. Mus., 38: 119– 133.
- 2 Michener, C. D. (1965) A classification of the bees of the Australian and South Pacific regions. Bull. Amer. Mus. Natl. Hist., 130: 1-362.
- 3 Blüthgen, P. (1926) Beiträge zur Kenntnis der indo-

malayischen Halictus- und Thrinchostoma-Arten (Hym., Apidae, Halictini). Zool. Jahrb., Syst., **51**: 375–698.

- 4 Blüthgen, P. (1931) Beiträge zur Kenntnis der indomalayischen Halictus- und Thrinchostoma-Arten (Hym., Apidae, Halictini). Zool. Jahrb., Syst., 61: 285-346.
- 5 Cockerell, T. D. A. (1937) African Bees of the Genera *Ceratina, Halictus* and *Megachile.* 254 pp., Brit. Mus., London.
- 6 Sakagami, S. F. (1989) Taxonomic notes on a Malesian bee Lasioglossum carinatum, the type species of the subgenus Ctenonomia, and its allies (Hymenoptera: Halictidae). J. Kansas entom. Soc., 62: 496– 510.

Addendum:

Ebmer (1987, Senkenbergia biol., 68: 84) transferred L. (N.) goluratum from Nesohalictus to Ctenonomia.